Quasiracemate Crystal Structures of Magainin 2 Derivatives Support the Functional Significance of the Phenylalanine Zipper Motif

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Supporting Information

ABSTRACT: Quasiracemic crystallography has been used to explore the significance of homochiral and heterochiral associations in a set of host-defense peptide derivatives. The previously reported racemic crystal structure of a magainin 2 derivative displayed a homochiral antiparallel dimer association featuring a “phenylalanine zipper” notable for the dual roles of phenylalanines in mediating dimerization and formation of an exposed hydrophobic swath. This motif is seen as well in two new quasiracemate crystals that contain the D form of the magainin 2 derivative along with an l-peptide in which one Ala has been replaced by a β-amino acid residue. This structural trend supports the hypothesis that the Phe zipper motif has functional significance.

Host-defense peptides (HDPs) constitute a ubiquitous component of the eukaryotic innate immune response to bacterial invasion.1 Collectively, these peptides exert multiple antimicrobial mechanisms of action, some that cause damage to bacterial cells and others that modulate immunological signaling in the host. A large HDP subset comprises relatively short and conformationally mobile amphiphilic peptides that are short and conformationally mobile are very rare.4

The bulk of the aromatic side chain cannot be buried within a coiled-coil interface; thus, six phenyl rings, from Phe5, Phe12 and Phe16 of each peptide, were arrayed along one face of each homochiral dimer in the crystal of 1. This type of aromatic ring arrangement has previously been designated a “phenylalanine zipper.”12 Dimers of this type had been detected via NMR under micellar conditions for peptides derived from magainin 2,13 although not for magainin 2 itself or for variant 1. These NMR

Figure 1. Peptides discussed in this paper. The structure of the β-amino acid residue designated ACPC is shown. Red A indicates a position at which the natural residue has been changed to alanine.

Magainin 2 is helical in the presence of vesicles,8 and the three residues altered in 1 are expected to enhance α-helical propensity.7 The crystal structure of 1 showed an α-helix that incorporated all residues except the C-terminal Ser, an observation consistent with previously described evidence of helicity in the presence of micelles.10 Of particular interest was an intimate association in the racemate crystal between two peptides with the same absolute configuration. These homochiral dimers featured an antiparallel coiled-coil-like interaction (Figure 2A; helices of the same color have the same configuration). Phe residues, unusual within canonical coiled-coil interfaces,11 played a prominent role in the homochiral association observed for 1. The bulk of the aromatic side chain cannot be buried within a coiled-coil interface; thus, six phenyl rings, from Phe5, Phe12 and Phe16 of each peptide, were arrayed along one face of each homochiral dimer in the crystal of 1. This type of aromatic ring arrangement has previously been designated a “phenylalanine zipper.”12 Dimers of this type had been detected via NMR under micellar conditions for peptides derived from magainin 2,13 although not for magainin 2 itself or for variant 1. These NMR

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Peptides $l$-2, $l$-3, and $l$-4 contain a single ACPC residue in place of Ala8, Ala13, or Ala18, respectively. Two quasiracemates, [($l$-2 + $d$-1)] and [(−$l$-3 + $d$-1)], were crystallized under the conditions that were successful for racemic $l$: 0.1 M sodium citrate tribasic, pH 5.6, 35% $v/v$ tert-butanol. Structure determination by molecular replacement and refinement using a new library of $d$-amino acid residues with canonical peptide bond restraints are described in the SI.

The [(−$l$-3 + $d$-1)] quasiracemate, refined at 2.2 Å resolution in space group $P2_12_12_1$, contains a pseudo-four-fold rotation axis that forms a tightly packed tetrameric quaternary arrangement (Figure 2B). If this were a true rotation axis, the space group would be $P4_2_12_1$. Each asymmetric unit contains two of these $l$-2 dimer + $d$-1 dimer “tetramers”, which differ slightly in the curvature of their $d$-1 peptides. As observed for [(−$l$-1 + $d$-1)], the C-terminal Ser residues of $l$-2 and $d$-1 could not be reliably modeled.

The asymmetric unit of the [(−$l$-3 + $d$-1)] quasiracemate contains four polypeptides, one $l$-3 dimer and one $d$-1 dimer that are related by a pseudoinversion center (Figure 2C). This structure, refined at 1.50 Å, can be viewed as pseudo-$P1$; however, the chemical distinction between the quasienantiomers required refinement in space group $P1$. The relationship between the dimers of the quasienantiomers in the [(−$l$-3 + $d$-1)] structure is pseudocentrosymmetric. (Packing representations and electron density maps for [(−$l$-2 + $d$-1)] and [(−$l$-3 + $d$-1)] are found in Figures 3S and 5S.)

In both of the quasiracemate structures, each component forms the type of homochiral coiled-coil dimer that was observed for the constituents of the racemic crystal of $l$. The Phe zipper motif is evident in each case (Figure 3). Backbone atom overlay data led to antibacterial evaluation of covalently dimerized magainin 2 derivatives. Our structure provided the first crystallographic support for the Phe zipper proposal in the magainin context. We proposed that the extended hydrophobic surface projected by the Phe zipper motif mediates association of the coiled-coil dimer with bacterial membranes.

In addition to the homochiral pairing in crystalline $l$, a heterochiral tetramer was observed in which the Phe zipper of an $l$-peptide dimer associates with the Phe zipper of a $d$-peptide dimer to create an aromatic-rich core (Figure 2A). We hypothesized that the heterochiral tetramer resulted from crystal packing effects, but this proposal raises the possibility that the homochiral dimer observed in the crystal of racemic $l$ is also a consequence of crystallization rather than a native mode of peptide assembly with potential relevance to antibacterial function. The new crystal structures described here support the hypothesis that the homochiral dimer observed in the crystal structure of racemic $l$ is functionally significant, whereas the heterochiral associations serve merely as crystallization contacts.

We undertook crystallization of quasiracemic mixtures that contain $d$-1 and a variant of $l$-1 to assess the consistency of homochiral and heterochiral associations. Quasiracemates, in which each component is slightly different from the enantiomer of the other, often display the crystallization advantages associated with true racemates. In the variants of $l$-1 we examined one Ala residue was replaced with the $\beta$-amino acid residue derived from $(S,S)$-trans-2-aminocyclopentanecarboxylic acid (ACPC; Figure 1). Previous work with oligomers that contain mixtures of $\alpha$- and $\beta$-amino acid residues (“$\alpha/\beta$-peptides”) has shown that the ACPC residue supports formation of a helical conformation strongly resembling the $\alpha$-helix.

Figure 2. Pairs of homochiral dimers form distinct heterochiral crystal packing arrangements in the structures of racemic $l$ (PDB 4MGP) and the two quasiracemates. In each case, the yellow molecules are $d$-1. (A) Racemic $l$; blue = $l$-1. (B) Quasiracemate [($l$-2 + $d$-1)]; green = $l$-2. (C) Quasiracemate [($l$-3 + $d$-1)]; red = $l$-3.

Figure 3. Overlay comparisons of peptide chains containing $l$-amino acid residues. Only the backbones (ribbon) and Phe side chains are shown. (A) $l$-1 (blue) from racemic $l$ (PDB 4MGP) vs $l$-2 (green) from the [($l$-2 + $d$-1)] quasiracemate. (B) $l$-1 (blue) from racemic $l$ (PDB 4MGP) vs $l$-3 (red) from the [($l$-3 + $d$-1)] quasiracemate.
rancem 1 (Figure 2B). However, the analogous tetrameric association in the \([\text{L-3} + \text{D-1}]\) quasiracemate is unique in that there is not a close packing of the phenylalanine zippers from each dimer (Figure 2C). The fact that the homochiral coiled-coil assembly, featuring the Phe zipper motif, is universal among the three data sets, involving five independent dimer structures, despite variations in the heterochiral associations, strongly supports the conclusion that the homochiral dimerization mode is relevant to a native interaction of magainin-family peptides, while the heterochiral associations reflect crystal packing forces that differ among the systems.

The two new ACPC-containing magainin 2 variants, L-2 and L-3, show similar or slightly superior levels of growth-inhibitory activity relative to L-1 against a small panel of bacteria (Table 1).

### Table 1. Minimum Inhibitory Concentration (MIC) for L- Peptides toward Four Bacterial Species

<table>
<thead>
<tr>
<th>Peptide</th>
<th>E. coli</th>
<th>B. subtilis</th>
<th>S. aureus</th>
<th>E. faecium</th>
</tr>
</thead>
<tbody>
<tr>
<td>L-1</td>
<td>12</td>
<td>6</td>
<td>50</td>
<td>50</td>
</tr>
<tr>
<td>L-2</td>
<td>12</td>
<td>6</td>
<td>12</td>
<td>12</td>
</tr>
<tr>
<td>L-3</td>
<td>12</td>
<td>6</td>
<td>12</td>
<td>25</td>
</tr>
<tr>
<td>L-4</td>
<td>12</td>
<td>6</td>
<td>12</td>
<td>12</td>
</tr>
</tbody>
</table>

However, both variants are somewhat more prone to cause lysis of human red blood cells (“hemolysis”) than is L-1 (Figure 4).

Our crystallographic data offer insights beyond those available via NMR, because the crystal structures reveal a mode of helix association that involves pairing of nonpolar surfaces but nevertheless leaves considerable nonpolar surface area exposed for interaction with a bacterial membrane. These observations are important in light of a proposal that helix dimerization would prevent hydrophobic surface exposure.

Our findings raise the possibility that quasiracemate crystallography will prove to be a general strategy for assessing polypeptide quaternary structure preferences. Specifically, exploration of multiple quasiracemates based on a given racemate structure appears to be a logical and efficient approach to obtaining sets of atomic-resolution data that collectively provide insights on noncovalent association preferences. This approach might be useful for other types of molecules as well.

### ASSOCIATED CONTENT

* Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b07206.

Experimental details for peptide synthesis and X-ray crystallography. Model coordinates and structure factors have been deposited in the Protein Data Bank as entries SCGN and SCGO (PDF).

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**Notes**

The authors declare no competing financial interest.

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